



Research Article

An integrated approach for herbal drugs to target GSK3B and its linkage with melanoma and type-2 diabetes

Dalia Alammari¹ and Nawal Helmi^{2*}

1 Department of Microbiology and Immunology, Faculty of Medicine, Ibn Sina National College, Jeddah, Saudi Arabia

2 Department of Biochemistry, College of Sciences, University of Jeddah, Jeddah, Saudi Arabia

* Correspondence: nmhelmi@uj.edu.sa (N.H.)

Citation: Alammari D and Helmi N. An integrated approach for herbal drugs to target GSK3B and its linkage with melanoma and type-2 diabetes. *Glob. Jour. Bas. Sci.* 2024, 1(2). 1-13.

Received: November 13, 2024 Revised: December 09, 2024 Accepted: December 21, 2024 Published: December 25, 2024

doi: 10.63454/jbs20000014

Abstract: Glycogen synthase kinase-3 (GSK3) is known to be among the busiest kinase in most cells which have more than 100 known substrates. The current understanding of GSK3 regulated mechanisms are emerging topics in the actions of GSK3, particularly its interactions with receptors and receptor-coupled signal transduction events, and differential actions and regulation of the two GSK3 isoforms, GSK3 α and GSK3 β . To understand the control of GSK3 control in terms of selective phosphorylation of the substrate and why does it appear evolutionarily favorable for GSK3 to assume such a large responsibility is potentially challenging main in case of human diseases. GSK3 must be particularly adaptable for incorporating new substrates into its repertoire, and we focused on it in unique way where it focused on the critical pathways and their major components which are common in the diseases type-2 diabetes and melanoma mediated by GSK3 β that may contribute to its capacity to fulfill its roles in multiple signaling pathways and finally, we have targeted these GSK3 associated proteins with the herbal drugs Diosmin and Fisetin. Here, we unravel the potential links between GSK3 β targets, melanoma, and type-2 diabetes and further presented pivotal herbal drugs and the targets for both the diseases mediated by GSK3 β mediates them.

Keywords: GSK3 β ; gene expression; herbal drugs; Diosmin and Fisetin; type-2 diabetes; and melanoma

1. Introduction

GSK3 is a widely expressed serine/threonine kinase that comes in two forms: GSK3 and GSK3 isoforms, both of which are active in the absence of phosphorylation by separate upstream kinases. GSK3 was first found as a glycogen synthesis regulator, but it's now implicated in a number of signaling pathways that govern a variety of activities. Hepatocellular carcinoma, cholangiocarcinoma, breast cancer, prostate cancer, leukaemia, and melanoma are all known to be linked to GSK3. It has been demonstrated to trigger apoptosis in some situations and prevent apoptosis in others, as well as to promote cancer development or limit tumor cell proliferation, indicating that various GSK3 modulators may target distinct targets). It also works on autophagy regulation, evaluating signaling pathways in neurodegenerative and liver illnesses, as well as oxidative stress and autophagic cell death, with an emphasis on liver injury. GSK3 is also thought to be a potential therapeutic target for natural and synthetic inhibitors in a variety of disorders, a regulator of mammalian ageing, and a link between metabolic changes in senescent cells and age-related diseases[1-4].

GSK3 inhibitors have previously been demonstrated to be effective in the treatment of SARS-CoV2. The phosphorylation of key serine residues in SARS-CoV2 nucleocapsid proteins by GSK3, which is necessary for viral replication, suggests that GSK3 may play a role in influencing innate and adaptive immune responses. GSK3 inhibition dampens hepatic and renal failure in animal models of hemorrhagic shock, according to studies, by upregulating antiinflammatory IL-10 and downregulating IL-12p40 and IL-6, a cytokine implicated in the cytokine release syndrome found in patients with severe SARS-CoV2. GSK3 inhibition lowers the systemic inflammatory response in models of sepsis and ischaemia/reperfusion injury through modulating the NF-kB-induced inflammatory response. SIR and the prevalent diffuse intravascular vascular coagulopathy found with SARS-CoV2 infection may be affected by these results. GSK3 inhibition lowered mRNA expression of IL-1, IL-6, and inducible NO synthase in a lipopolysaccharide (LPS)-mediated inflammatory model (iNOS)[3, 5-7]. Diabetes is a high-risk illness that is connected to heart disease and stroke. According to the WHO, diabetes will affect 511 million people by 2030. Many targets have been found in the hunt for new targets for type-2 diabetes, including GSK3, Dipeptidyl Peptidase, PPAR, Glucosidase, Amylase, GLP-1, and SGLT. Among the targets, GSK3 has been found to be an effective therapy for diabetes. GSK3 is a glycogen synthesis regulating enzyme in glycogen metabolism (glycogenesis). It catalyzes the creation of 1,4-glycosidic linkages in a linear, nonbranched molecule. GSK3 is a family of semi-conservative multifunctional serine/threonine kinase enzymes that comprises two isoenzymes with different N- and C-terminal sequences. Previous research has also revealed that there are distinct routes for diabetes and melanoma, as well as how GSK3 alteration may influence both. A mutation in the GSK3 complex causes diabetes. Synthetic and natural scaffolds affect GSK3, which has led to its optimization for the development of GSK3 inhibitors[8-14].

There are a number of previous research work focused on the role of GSK3 signaling in melanoma carcinogenesis. According to immunohistochemistry, GSK3 was shown to be focally expressed in the invasive portions of 12 and 33 percent of primary and metastatic melanomas, respectively. GSK3 inhibitors and siRNA knockdown of GSK3 were reported to reduce the motile behavior of melanoma cells in scratch wound, three-dimensional collagenimplanted spheroid, and modified Boyden chamber tests. GSK3 signaling inhibition was connected to lower transcription factor expression Slug and was shown to restrict N-cadherin expression at the messenger RNA and protein levels. Forced overexpression of N-cadherin reversed the impact of pharmacological and genetic GSK3 signaling ablation, which inhibited melanoma cells from adhering to endothelial cells and fibroblasts and preventing transendothelial migration. The ability of GSK3 inhibitors and siRNA knockdown to prevent FAK phosphorylation and increase the number of focal adhesions shows that GSK3 signaling plays another function in invasion. To summarize, we have discovered a previously undiscovered role for GSK3 in changing the motile and invasive behavior of melanoma cells via N-cadherin and FAK, to our knowledge. According to this study, inhibiting GSK3 may have therapeutic relevance in certain kinds of melanoma[15, 16].

With over 100 identified substrates to deal with, GSK3 may be the busiest kinase in most cells. How does GSK3 preserve its ability to selectively phosphorylate each substrate, and why was it advantageous for GSK3 to take on such a big role in evolution? GSK3 must be very versatile to add new substrates to its repertoire, and we examine the unique characteristics of GSK3 that may contribute to its ability to play diverse roles in signaling circuits. The regulatory mechanisms governing GSK3 (primarily post-translational modifications, substrate priming, cellular trafficking, and protein complexes) have previously been discussed, as well as how each of these regulatory mechanisms contributes to GSK3's ability to select which substrates to phosphorylate, and how these mechanisms may have contributed to its adaptability as new substrates evolved. Most researchers' current focus is on the periphery of their understanding of the mechanisms regulating GSK3 and emerging topics in GSK3's actions, particularly its interactions with receptors and receptor-coupled signal transduction events, as well as the differences in the actions and regulation of the two GSK3 isoforms, GSK3 and GSK3. GSK3 is also known for its participation in a variety of illnesses, including mental and neurological disorders, inflammatory diseases, cancer, and others. We have presented the therapeutic potential of targeting GSK3. We have concentrated on known interactors based on the protein-protein interaction (PPI) network database, with a particular focus on the link with melanoma and prospective herbal medicines Diosmin and Fisetin to target. We've also looked at the link between GSK3 and T2D, although our main focus is still on GSK3 and melanoma[1-3, 7, 17-31].

2. Methods

Here, the main goal was to study the linkage of GSK3ß in case of mainly melanoma and T2D in terms of the gene expression and their functional impact, for which gene expression datasets were collected from Gene Expression (GEO). Omnibus The primary source data is GSE41662 of our (https://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE41662[32]), TCGA database for melanoma and GSE121 (https://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE121[33]). Since, we were mainly interested in understanding the potential linkage of GSK3ß with melanoma and also with T2D and the difference between gene expression pattern. We used GEO2R to generate a list of DEGs by dividing samples into two groups: control and infected, and uninfected and infected, and then calculating fold changes and p-values. The thresholds for fold changes and p-values used to select genes as differentially expressed were +/- 2 and 0.05, respectively. We used the KEGG database for pathway research and developed our own algorithm for pathway and network analysis. KEGG Pathways was used for associated pathways with GSK3β, Diosmin and Fisetin[25, 34-37] and their interactors (STITCH database) followed by the protein classes (SwissTargetPrediction), and (c) Fisetin and its interactors (STITCH database) followed by the protein classes (SwissTargetPrediction)[20, 23, 26, 28, 38-40].

MATLAB algorithms, such as mattest, have been used for statistical analysis and differential gene expression prediction. The KEGG database and proprietary code for pathway and network analysis were used for pathway analysis. Throughout the project, FunCoup2.0 was utilised to generate the DEGs networks, and Cytoscape was utilised to visualise the networks. Four sorts of functional couplings or linkages, including protein complexes, physical interactions

between proteins, metabolic processes, and signalling pathways, are predicted by FunCoup. The majority of our computations and coding have been done using MATLAB[20, 23, 26-30, 34, 39-45].

After researching the DEGs and enriched pathways, the next step was to understand the network and the connections between the genes inside the DEG network. This was done using the FunCoup2.0 database, and the network was viewed using Cytoscape. MATLAB code was also used for figure plotting and analysis. MATLAB was used to generate the scripts for network level analysis, including how many connections each gene has and how many pathways each gene belongs to. MATLAB routines were also used to display the data. Swissmodel will be used to predict the 3D protein structure of the selected genes, and Swissdock will be utilised for docking[20, 34, 41, 45, 46].

3. Results

3.1. GSK3β controls a number of critical biological pathways and functions: Here, we have mapped out all the

pathways associated where GSK3β is the direct component (Figure 1a). Among these pathways, there are a number of pathways which directly controls human diseases mainly cancers and infectious diseases or the disease associated with immune system. The critical pathways such as Wnt signaling, cell cycle chemokine signaling, BCR/TCR signaling, PI3K-AKT signaling, and insulin signaling appear common to both the diseases type-2 diabetes and melanoma. The other pathways are well-defined in case of different types of cancers includina melanoma (melanogenesis, pathways in cancer, colorectal cancer, endometrial cancer, prostate cancer) (Figure 1a). Based on these preliminary outcome, it appears that GSK3^β –focused study could lead to potential therapeutic findings so we have applied in-silico approach where used two herbal drugs Diosmin and Fisetin to target the potential pathway(s) components to understanding the binding possibility of the two selected drugs (Diosmin and Fisetin). As an initial step, we have evaluated the binding targets of these two herbal drugs by using SwissTargetPrediction and classified the target proteins (Figure 1b and 1c). As compared to Fisetin, Diosmin shows much larger number of interactors. In terms of protein classes also both these drugs have different protein

classes in dominance. In case of Diosmin, lyase and family A, G-protein-coupled receptor, secreted proteins, and enzyme are 26.7%, 26.7%, 13.3%, and 13.3%, respectively. Enzyme (33.3%), kinase (26.7%), and oxidoreductase (13.3%) are dominantly present as interactors of Fisetin (Figure 1b and 1c).

3.2. Genes and pathways shared between the GSK3ß interactors, type-2 diabetes, and melanoma: In the next step, we have performed the mapping of overall GSK3β-associated genes, DEGs of melanoma, and DEGs of T2D and pathways enrichment analysis for the respective genes lists and finally performed comparative analysis of all the three

lists in terms of genes and pathways (Figure 1d and 1e). From the comparative study of the three genes lists, we observe that there were 10 genes shared between GSK3β and melanoma, 20 genes shared between GSK3β and T2D while melanoma and T2D does not show any shared gene and also we could not see even a single gene common between all three lists. So, we could conclude that there could be shared genes between GSK3β and melanoma and GSK3β and T2D which could act as bridge between them while there is no common gene between T2D and melanoma (Figure 1d). Similar to gene comparison, we have also performed the comparative study of the biological pathways and the functions for the three genes lists and we observe that there is only one pathway which is common between all the three lists (Figure 1e and Table 1). There were seven pathways common between

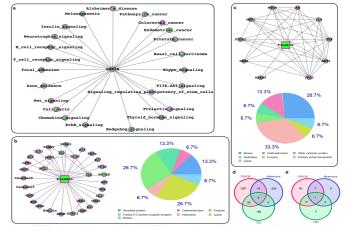


Figure 1. Basic analysis of GSK3β, Mimosin, and Fisetin. (a) KEGG Pathways associated with GSK3β, (b) Mimosin and its interactors (STITCH database) followed by the protein classes (SwissTargetPrediction), and (c) Fisetin and its interactors (STITCH database[51]) followed by the protein classes (SwissTargetPrediction).

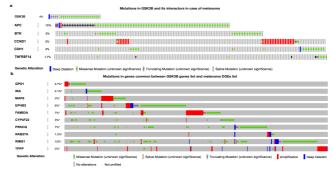


Figure 2. (a) Mutations in GSK3B and its interactors in case of melanoma and (b) Mutations in genes common between GSK3B genes list and melanoma DGEs list.

GSK3β and melanoma and 17 pathways common between GSK3β and T2D and there was not a single pathway shared between T2D and melanoma. Cytokine-cytokine receptor interaction appears to be commonly enriched for GSK3β genes list, melanoma, and T2D. Purine metabolism, starch and sucrose metabolism, retinol metabolism, metabolism of xenobiotics by cytochrome P450, NF--kB signaling, Toll-like receptor signaling, adipocytokine signaling between GSK3B and melanoma (Table 1). ErbB signaling, cAMP signaling, FoxO signaling, Phagosome, PI3K-Akt signaling, apoptosis, adrenergic signaling in cardiomyocytes, wnt signaling, osteoclast differentiation, hippo signaling, focal adhesion, pathways regulating pluripotency of stem cells, platelet activation, Jak-STAT signaling, regulation of actin cytoskeleton, insulin signaling pathway, oxytocin signaling between GSK3β and T2D (Table 1).

 Table 1. Enriched pathways for GSK3B, melanoma, and type-2 diabetes.

 Enriched pathways for GSK3B-associated genes

Enriched pathways	p-values
KEGG_04010_MAPK_signaling	5.42E-20
KEGG_04012_ErbB_signaling	5.42E-20
KEGG_04014_Ras_signaling	5.42E-20
KEGG_04015_Rap1_signaling	5.42E-20
KEGG_04020_Calcium_signaling	5.42E-20
KEGG_04022_cGMP-PKG_signaling	5.42E-20
KEGG_04024_cAMP_signaling	5.42E-20
KEGG_04066_HIF-1_signaling	5.42E-20
KEGG_04068_FoxO_signaling	5.42E-20
KEGG_04071_Sphingolipid_signaling	5.42E-20
KEGG_04072_Phospholipase_D_signaling	5.42E-20
KEGG_04110_Cell_cycle	5.42E-20
KEGG_04114_Oocyte_meiosis	5.42E-20
KEGG_04120_Ubiquitin_mediated_proteolysis	5.42E-20
KEGG_04151_PI3K-Akt_signaling	5.42E-20
KEGG_04152_AMPK_signaling	5.42E-20
KEGG_04211_Longevity_regulating_pathway	5.42E-20
KEGG_04261_Adrenergic_signaling_in_cardiomyocytes	5.42E-20
KEGG_04310_Wnt_signaling	5.42E-20
KEGG_04360_Axon_guidance	5.42E-20
KEGG_04370_VEGF_signaling	5.42E-20
KEGG_04371_Apelin_signaling	5.42E-20
KEGG_04380_Osteoclast_differentiation	5.42E-20
KEGG_04392_Hippo_Signaling	5.42E-20
KEGG_04510_Focal_adhesion	5.42E-20
KEGG_04520_Adherens_junction	5.42E-20
KEGG_04530_Tight_junction	5.42E-20
KEGG_04540_Gap_junction	5.42E-20
KEGG_04550_Signalings_regulating_pluripotency_of_stem_cells	5.42E-20
KEGG_04611_Platelet_activation	5.42E-20
KEGG_04650_Natural_killer_cell_mediated_cytotoxicity	5.42E-20
KEGG_04660_T_cell_receptor_signaling	5.42E-20
KEGG_04662_B_cell_receptor_signaling	5.42E-20
KEGG_04664_Fc_epsilon_RI_signaling	5.42E-20
KEGG_04666_Fc_gamma_R-mediated_phagocytosis	5.42E-20

KEGG_04668_TNF_signaling	5.42E-20
KEGG_04713_Circadian_entrainment	5.42E-20
KEGG_04720_Long-term_potentiation	5.42E-20
KEGG_04721_Synaptic_vesicle_cycle	5.42E-20
KEGG_04722_Neurotrophin_signaling	5.42E-20
KEGG_04750_Inflammatory_mediator_regulation_of_TRP_channels	5.42E-20
KEGG_04810_Regulation_of_actin_cytoskeleton	5.42E-20
KEGG_04910_Insulin_signaling	5.42E-20
KEGG_04912_GnRH_signaling	5.42E-20
KEGG_04914_Progesterone-mediated_oocyte_maturation	5.42E-20
KEGG_04915_Estrogen_signaling	5.42E-20
KEGG_04916_Melanogenesis	5.42E-20
KEGG_04917_Prolactin_signaling	5.42E-20
KEGG_04919_Thyroid_hormone_signaling	5.42E-20
KEGG_04920_Adipocytokine_signaling	5.42E-20
KEGG_04921_Oxytocin_signaling	5.42E-20
KEGG_04145_Phagosome	4.11E-19
KEGG_04270_Vascular_smooth_muscle_contraction	4.11E-19
KEGG_04723_Retrograde_endocannabinoid_signaling	4.11E-19
KEGG_04730_Long-term_depression	4.11E-19
KEGG_04210_Apoptosis	8.22E-18
KEGG_04620_Toll-like_receptor_signaling	8.22E-18
KEGG_03010_Ribosome	1.56E-16
KEGG_04670_Leukocyte_transendothelial_migration	1.56E-16
KEGG_04144_Endocytosis	2.81E-15
KEGG_04918_Thyroid_hormone_synthesis	2.81E-15
KEGG_00230_Purine_metabolism	4.78E-14
KEGG_04141_Protein_processing_in_endoplasmic_reticulum	4.78E-14
KEGG_00020_Citrate_cycle_(TCA_cycle)	7.65E-13
KEGG_00030_Pentose_phosphate_pathway	7.65E-13
KEGG_00051_Fructose_and_mannose_metabolism	7.65E-13
KEGG_00190_Oxidative_phosphorylation	7.65E-13
KEGG_04150_mTOR_signaling	7.65E-13
KEGG_00500_Starch_and_sucrose_metabolism	1.15E-11
KEGG_00620_Pyruvate_metabolism	1.15E-11
KEGG_04340_Hedgehog_signaling	1.15E-11
KEGG_04350_TGF-beta_signaling	1.15E-11
KEGG_04630_Jak-STAT_signaling	1.15E-11
KEGG_03013_RNA_transport	1.61E-10
KEGG_04622_RIG-I-like_receptor_signaling	2.09E-09
KEGG_00520_Amino_sugar_and_nucleotide_sugar_metabolism	2.51E-08
KEGG 04064 NF-kappa B signaling	2.51E-08

KEGG_04070_Phosphatidylinositol_signaling_system	2.51E-08
KEGG_04924_Renin_secretion	2.51E-08
KEGG_00052_Galactose_metabolism	2.76E-07
KEGG_00562_Inositol_phosphate_metabolism	2.76E-07
KEGG_03320_PPAR_signaling	2.76E-07
KEGG_04140_Regulation_of_autophagy	2.76E-07
KEGG_04740_Olfactory_transduction	2.76E-07
KEGG_00071_Fatty_acid_metabolism	2.76E-06
KEGG_00240_Pyrimidine_metabolism	2.76E-06
KEGG_04060_Cytokine-cytokine_receptor_interaction	2.76E-06
KEGG_04115_p53_signaling	2.76E-06
KEGG_04960_Aldosterone-regulated_sodium_reabsorption	2.76E-06
KEGG_00564_Glycerophospholipid_metabolism	2.48E-05
KEGG_04130_SNARE_interactions_in_vesicular_transport	2.48E-05
KEGG_00710_Carbon_fixation_in_photosynthetic_organisms	4.18E-05
KEGG_04962_Vasopressin-regulated_water_reabsorption	1.88E-04
KEGG_00590_Arachidonic_acid_metabolism	1.98E-04
KEGG_00970_Aminoacyl-tRNA_biosynthesis	1.98E-04
KEGG_03040_Spliceosome	1.98E-04
KEGG_03050_Proteasome	1.98E-04
KEGG_04146_Peroxisome	1.98E-04
KEGG_04330_Notch_signaling	1.98E-04
KEGG_04621_NOD-like_receptor_signaling	1.98E-04
KEGG_00260_Glycineserine_and_threonine_metabolism	1.39E-03
KEGG_00561_Glycerolipid_metabolism	1.39E-03
KEGG_00980_Metabolism_of_xenobiotics_by_cytochrome_P450	1.39E-03
KEGG_04742_Taste_transduction	1.39E-03
KEGG_04913_Ovarian_steroidogenesis	1.39E-03
KEGG_00330_Arginine_and_proline_metabolism	8.33E-03
KEGG_00480_Glutathione_metabolism	8.33E-03
KEGG_00650_Butanoate_metabolism	8.33E-03
KEGG_03420_Nucleotide_excision_repair	8.33E-03
KEGG_00250_Alanineaspartate_and_glutamate_metabolism	4.17E-02
KEGG_00270_Cysteine_and_methionine_metabolism	4.17E-02
KEGG_00280_Valineleucine_and_isoleucine_degradation	4.17E-02
KEGG_00340_Histidine_metabolism	4.17E-02
KEGG_00380_Tryptophan_metabolism	4.17E-02
KEGG_00450_Selenoamino_acid_metabolism	4.17E-02
KEGG_00640_Propanoate_metabolism	4.17E-02
KEGG_00830_Retinol_metabolism	4.17E-02
KEGG_04514_Cell_adhesion_molecules_(CAMs)	4.17E-02
KEGG 04612 Antigen processing and presentation	4.17E-02

Enriched pathways for GSK3B-associated genes Melanon	na
KEGG_00830_Retinol_metabolism	2.09E-09
KEGG_00983_Drug_metabolismother_enzymes	2.09E-09
KEGG_00980_Metabolism_of_xenobiotics_by_cytochrome_P450	2.51E-08
KEGG_00982_Drug_metabolismcytochrome_P450	2.51E-08
KEGG_00140_Steroid_hormone_biosynthesis	2.76E-07
KEGG_00500_Starch_and_sucrose_metabolism	2.76E-06
KEGG_00860_Porphyrin_and_chlorophyll_metabolism	2.76E-06
KEGG_00040_Pentose_and_glucuronate_interconversions	2.09E-05
KEGG_04060_Cytokine-cytokine_receptor_interaction	2.48E-05
KEGG_04064_NF-kappa_B_signaling	1.39E-03
KEGG_04620_Toll-like_receptor_signaling	8.33E-03
KEGG_00230_Purine_metabolism	4.17E-02
KEGG_04920_Adipocytokine_signaling	4.17E-02

Enriched pathways for GSK3B-associated genes Melanoma

Enriched pathways for GSK3B-associated genes T2D

KEGG_04151_PI3K-Akt_signaling	2.51E-08
KEGG_04810_Regulation_of_actin_cytoskeleton	2.76E-06
KEGG_04510_Focal_adhesion	2.48E-05
KEGG_04024_cAMP_signaling	8.33E-03
KEGG_04068_FoxO_signaling	8.33E-03
KEGG_04145_Phagosome	8.33E-03
KEGG_04261_Adrenergic_signaling_in_cardiomyocytes	8.33E-03
KEGG_04310_Wnt_signaling	8.33E-03
KEGG_04380_Osteoclast_differentiation	8.33E-03
KEGG_04392_Hippo_Signaling	8.33E-03
KEGG_04611_Platelet_activation	8.33E-03
KEGG_04910_Insulin_signaling	8.33E-03
KEGG_00310_Lysine_degradation	4.17E-02
KEGG_04012_ErbB_signaling	4.17E-02
KEGG_04060_Cytokine-cytokine_receptor_interaction	4.17E-02
KEGG_04210_Apoptosis	4.17E-02
KEGG_04512_ECM-receptor_interaction	4.17E-02
KEGG_04550_Signalings_regulating_pluripotency_of_stem_cells	4.17E-02
KEGG_04630_Jak-STAT_signaling	4.17E-02
KEGG_04921_Oxytocin_signaling	4.17E-02

3.3. Genes and pathways shared between the GSK3β interactors, type-2 diabetes, and melanoma: Now, we have analyzed the genes which were common between the three lists and observe that GPD1, INA, MAP6, EPHB2, FAM83A, CYP4F22, PRKCQ, RAB27A, RIMS1, and TPPP were common between GSK3β genes list and melanoma DGEs list. ACVR1B, APC, ARHGAP44, BTK, CAMK2B, CCND1, CDH1, EIF5B, GAP43, GLIPR1, OGT, PAK2, PPP1CA, PPP2R5E, PRKAR2A, RAB3GAP2, STX7, TNFRSF14, TRIM3, and TUBA1B were common between GSK3β genes list and the T2D DEGs list (Table 1).

We have further explored the genes common between GSK3β genes list and melanoma DGEs list, genes common between GSK3β genes list and the T2D DEGs list, Diosmin and Fisetin interactors. For this purpose, we have used the

TCGA database mainly to investigate the potentials of these genes in melanoma. Here, we have explored the mutations in the inferred genes of GSK3 β interactors, GSK3 β genes list and melanoma DGEs list, genes common between GSK3 β genes list and the T2D DEGs list, and the mutations in inferred genes of Diosmin and Fisetin interactors in case of melanoma. We observe that in case of inferred genes of GSK3 β interactors, APC appear highly mutated in case of melanoma, BTK mutated in 5%, CDH1 mutated in 4%, and TNFSF14 mutated in 1.7% of total selected melanoma samples (Figure 2a).

In case of the genes common between GSK3β genes list and melanoma DGEs list, much higher number of melanoma patients shows mutations in RIMS1 (12%), followed by EPHB2 (9%), FAM83A (7%), and PRKCQ (7%). MAP6, TPPP, CYP4F22, INA, and RAB27A appear mutated in \leq 3% of the total selected melanoma patients (Figure 2b). Moreover, the mutations have been deeply explored to understand the mutation regions (domains/motifs) and the type of mutations (Figure 3). None of these genes display driver and SV/fusion mutation while VUS (variant of uncertain (or unknown) significance) which is a genetic variant that has been identified through genetic testing but whose significance to the function or health of an organism is not known and missense mutations are dominantly present. In case of the genes common between GSK3ß genes list and T2D DGEs list, APC and ARHGAP44 appear mutated in 11% and 8% of the total melanoma patients, respectively. BTK, CCND1, and RAB3GAP2 were

mutated in 5%, 5%, and 4% of the total

respectively and the rest of the genes were mutated in less than 4% of the melanoma patients (Figure 4). Similarly, we have also explored the types of mutations and their locations further where we observe opposite to

the genes common

melanoma mainly in

of

list

between

genes

case

patients,

GSK3_β

and

driver

melanoma

ACVR18		Barrier 1997 1997					
	1.5%*						
APC	11%*	··· •					
ARHGAP44	8%*	H 4 4					
BTK	5%*	A CONTRACTOR OF A			_		
CAMK2B	3%*			1.10	-		
CCND1	6%*			1.00		-	
CDH1	3%*	1			1.1	-	
EIFSB	1.0%*	00000-0010-0		0 0		1	
GAP43	3%*	10 III - III				10.0	
GLIPR1	2.4%*	10101-011-014				1	
OGT	3%*	000100-0010-0-0		- P		10 C 10 P	
PAK2	2.7%*	10100-0010-010		1			
PPP1CA	2.9%*	X				10 1	k
PPP2R5E	1.0%*	100 C 100 C 101 C 101 C					-
PRKAR2A	1.6%	NOTE: 014 014		11		1	1
RAB3GAP2	4%*	100 K 000 0 1				100 00000	-
STX7	1.216*	No					
TNFRSF14	2.1%	1	1.1.1		0.0		1.1
TRIM3	2.5%*	AUGUST - 101-0-1				1.0	1
TUBA1B	1.8%*					1.0	1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1
		Gond	tic Alte	oratio			
_							
Inframe Mutation	(unk	nown significi	ance)	Mis	sense Mul	ation (putati	ve driver)
Splice Mutation (unkno	own significar	nce)	Trun	cating Mut	ation (putativ	/e driver)
Structural Variant	(unk	nown signific	ance)	Am	plification	Deep [Deletion
-		-					
Missense Mutati	on (u	nknown signifi	cance)	Sp	lice Mutatio	on (putative o	lriver)
Truncating Mutat	ion (u	nknown signif	icance)	St	ructural Va	riant (putativ	e driver)
No alterations	Not	profiled					
I no anerations	1400	promed					

Figure 4. Mutations in the genes common between GSK3 β genes list and the T2D DEGs list with their respective 3D protein structures.

mutations which shows driver mutations are dominantly present in several genes (APC (64), CCND1 (5), CDH1 (7), and STX7 (1)) in case of the genes GSK3 β genes list and T2D DEGs list (Figures 5--6).

Finally, we have explored the mutations in the inferred genes of Diosmin and Fisetin interactors in case of melanoma patient samples. Here, there are a number of genes which were highly mutated in melanoma patients. CDKN2A appear mutated in 32% of the total patients mostly were deep

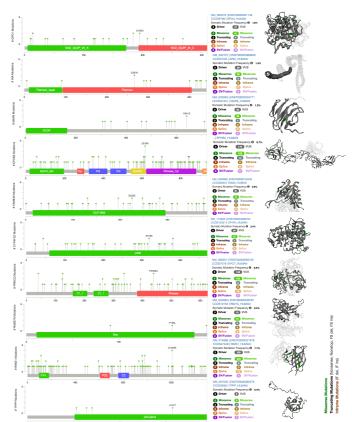


Figure 3. Mutations in the genes common between GSK3B genes list and melanoma DGEs list.

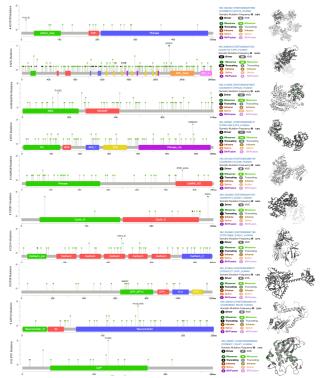


Figure 5. Mutations in the genes common between GSK3B genes list and T2D DGEs list.

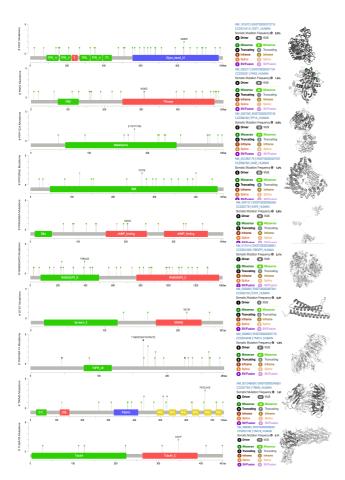


Figure 6. Mutations in the genes common between GSK3B genes list and T2D DGEs list.

GSK3 is a widely expressed serine/threonine kinase that comes in two forms: GSK3 and GSK3 isoforms, both of which are active in the absence of phosphorylation by separate upstream kinases. GSK3

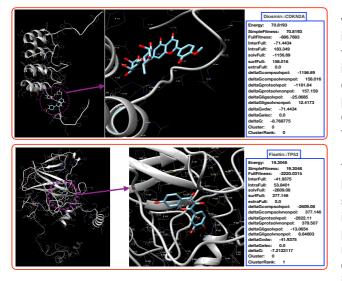


Figure 8. Docking of CDKN2A with Diosmin and TP53 with Fisetin.

deletions (Figure 7) and this gene is inferred for Diosmin interactor. TP3 and MTOR of Fisetin was mutated in 18% and 9% of the patients, respectively. Moreover, CDKN2A and TP53 were docked with Diosmin and Fisetin, respectively and we observe that Diosmin has much higher binding affinity (delta G = -8.76kcal/mol) compared to TP53 docked with Fisetin (delta G = -7.21kcal/mol) while the delta G of Fisetin with TP53 is also quite significant (Figure 8). This result leads to the conclusion that for such critical proteins CDKN2A and TP53, Diosmin and Fisetin could be the pivotal drugs to target melanoma.

4. Dicussion: We focused on known interactors based on the PPI network database, with a specific focus on the relationship with melanoma and possible herbal drugs to target, in order to debate and show the potentials of GSK3 in the case of melanoma. Although our major emphasis is still GSK3 and melanoma, we've looked at the link between GSK3 and T2D. Such research takes a novel approach to GSK3 and its link to melanoma, followed by T2D, which is highly unusual in terms of its nobility.

				Diosmin genes mutation			
		1.4%*					
		1.0%*					
		6%*					
		2.6%*					
		6%*				11	
		2256*		,			
		0.9%*		-			
		3876*	beitt tild beren i itt beite beiter wither biller				
		2.7%*					
		6757		all and a second			A REAL PROPERTY AND A REAL
		2.275					
		4757					
		2.875*	and the second				
		975*					ALTERNATION AND ADDRESS OF ADDRESS OF
		2.3%					
		879.7	a contract of the second se				
		2,1197	and the second		and the second second		A CONTRACTOR OF A
		2767	and the second se	111 1 1 1 1 1		11.00	I I I I I I I I I I I I I I I I I I I
		2.735			-12-1400 - 14-02-02-02000		
		2.1%*					1
	1	37947	ALC: 10 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1	0.000 0 1 0 0	10.100 10.000	41 x	time was successive
	Ξ	3.7%*	••• ••• ••• ••• ••• ••• ••• •••		1110 1010	1.00	
	Ξ	3.1167	R	10.0400 1 1 10.000	11100 10 00 0000	1 C C C C C C C C C C C C C C C C C C C	11000 00100 100000 01100
	Ξ	675*	(MAX 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1				1000 00100 100000 00100
	Ξ	3.3%*				1 1 1	
ion			E Splice Mutation (putative driver)		annae Mutation (putative driver) Eng Mutation (putative driver) pification	Missense Mulation (unknown significance Truncating Mulation (unknown significance) No attentions - Not profiled	
					on in melanoma		
		2.0%	•				
	:	2.8% 2.1%					
	÷	2.8%* 2.1%* 4%*					
	1	2.8% 2.1% 4% 1.4%					_
	÷	2.8%* 2.1%* 4%*					
		2,8% 2,1% 4% 1,4% 0,7%					
	1	2.8% ² 2.1% ² 4% ² 1.4% ² 6.7% ²					
		2.8% 2.4% 4% 4% 6.7% 6% 8% 8%					
		2.8%* 2.1%* 4%* 1.4%* 0.7%* 6%*					

Figure 7. Mutations in the inferred genes of Diosmin and Fisetin interactors.

was first found as a glycogen synthesis regulator, but it's now implicated in a number of signaling pathways that govern a variety of activities. It is involved in a variety of disorders, including cancer and T2D. The active form of GSK3 has been demonstrated to trigger apoptosis in some situations and prevent apoptosis in others, as well as to promote cancer development or limit tumor cell proliferation, implying that various GSK3 modulators may target distinct targets[2, 4, 5, 9, 18, 19, 22, 47].

The involvement of GSK3 β , a multifunctional serine/threonine kinase found in all eukaryotes, in insulin-dependent glycogen synthesis was initially discovered. GSK3 has been shown to be involved in a number of cellular functions, including proliferation, differentiation, motility, and survival. GSK3 β malfunction has been associated to non-insulin-dependent diabetes, cardiovascular disease, many neurodegenerative illnesses, and bipolar disorder, among other things. As a result, GSK3 β inhibitors' medicinal potential has been a prominent research focus. However, GSK3 β has a role in tumor growth

and neoplastic transformation. The role of GSK3 in carcinogenesis and cancer progression is still being contested; it may operate as a "tumor suppressor" in certain tumors while encouraging growth and development in others. GSK3β

ADAMI ARTI CDK8 IL4 MMP1 MTOR PTG52 TNF TP53 controls drug sensitivity and resistance in cancer treatment. As a result, while GSK3β is a prospective therapeutic target for a range of human illnesses, its impact on cancer development and therapy must be carefully examined. This gene produces a serine-threonine kinase that belongs to the glycogen synthase kinase subfamily. It is a negative regulator of glucose homeostasis and has a role in energy metabolism, inflammation, ER-stress, mitochondrial dysfunction, and apoptotic pathways. Mutations in this gene have been associated to Parkinson's disease and Alzheimer's disease[1, 48-55].

Table 2. Mutual exclusivity.

			Gene A	Gene B					
			Not Gene	Not Gene		Log2 Odds			
Gene A	Gene B	Neither	В	А	Both	Ratio	p-Value	q-Value	Tendency
									Co-
PRKCQ	RIMS1	1262	62	145	49	2.782	<0.001	<0.001	occurrence
									Co-
CYP4F22	RIMS1	1298	26	165	29	>3	<0.001	<0.001	occurrence
									Co-
CYP4F22	PRKCQ	1371	36	92	19	2.975	<0.001	<0.001	occurrence
									Co-
INA	CYP4F22	1443	20	45	10	>3	<0.001	<0.001	occurrence
									Co-
EPHB2	RIMS1	1215	109	153	41	1.579	<0.001	<0.001	occurrence
									Co-
GPD1	RIMS1	1298	26	175	19	2.438	<0.001	<0.001	occurrence
									Co-
EPHB2	PRKCQ	1285	122	83	28	1.829	<0.001	<0.001	occurrence
									Co-
GPD1	CYP4F22	1427	36	46	9	2.955	<0.001	<0.001	occurrence
									Co-
RAB27A	TPPP	1448	16	48	6	>3	<0.001	<0.001	occurrence
									Co-
INA	RIMS1	1306	18	182	12	2.258	<0.001	<0.001	occurrence
									Co-
INA	PRKCQ	1386	21	102	9	2.542	<0.001	<0.001	occurrence
		10.10			10				Co-
INA	EPHB2	1348	20	140	10	2.267	<0.001	0.001	occurrence
FRURA	0)/04500	4007	100	44		4 700	-0.001	0.000	Co-
EPHB2	CYP4F22	1327	136	41	14	1.736	<0.001	0.002	occurrence
FAM83A	DIMO	1040	70	170	04	1 470	0.004	0.005	Co-
FAIVIOJA	RIMS1	1246	78	170	24	1.173	0.001	0.005	occurrence
FAM83A	TPPP	1372	92	44	10	1.761	0.002	0.007	Co-
FAIVIOJA		13/2	32	44	10	1.701	0.002	0.007	occurrence
FAM83A	PRKCQ	1320	87	96	15	1 245	0.005	0.015	Co-
FAIVI83A	PRACQ	1320	87	96	15	1.245	0.005	0.015	occurrence

									Co-
GPD1	FAM83A	1379	37	94	8	1.665	0.008	0.021	occurrence
									Co-
EPHB2	FAM83A	1284	132	84	18	1.06	0.009	0.021	occurrence
									Co-
FAM83A	CYP4F22	1370	93	46	9	1.527	0.009	0.022	occurrence
									Co-
GPD1	EPHB2	1333	35	140	10	1.444	0.01	0.022	occurrence
									Co-
MAP6	CYP4F22	1414	49	49	6	1.821	0.013	0.027	occurrence
									Co-
MAP6	PRKCQ	1361	46	102	9	1.384	0.016	0.032	occurrence
									Co-
MAP6	RIMS1	1282	42	181	13	1.132	0.017	0.034	occurrence

T2D is a non-communicable disease that causes blood glucose levels to remain consistently high outside of the normal range. Diabetes and insulin resistance are the world's major causes of illness and death in humans. A variety of enzymes and hormones have a role in this illness, the most significant of which are GSK3, a key enzyme, and insulin, a critical hormone. GSK3, the main enzyme, controls and influences cellular shape, growth, motility, and apoptosis by a number of mechanisms, including phosphorylation, protein complex formation, and other cellular distribution. GSK3 enzyme dysfunction can cause a number of disorders, including insulin resistance and diabetes, as well as neurological ailments including Alzheimer's disease and cancer. Fluoroquinolones are the most common class of drugs that cause dysglycemia through interacting with the GSK3 enzyme. As a result, understanding GSK3's actions and processes is crucial, notably its function in glucose homeostasis via effects on glycogen synthase.

Obesity and T2D have both been related to an increased cancer risk and are becoming more prevalent. Insulin resistance and dyslipidemia are metabolic diseases that have been associated to obesity and type 2 diabetes, as well as the obesity-cancer relationship. Increased insulin/IGF-1 signaling, lipid and glucose uptake and metabolism, changes in cytokine, chemokine, and adipokine profiles, as well as changes in the adipose tissue immediately adjacent to cancer sites, have all been hypothesized as pathways associating obesity and diabetes with cancer progression.

5. Conclusions: Here, we have performed GSK3ß based study with the potential focus on melanoma and additional focus on type-2 diabetes and the two herbal drugs in integration with GSK3ß oriented study in melanoma. This study leads to the conclusion that. GSK3β controls a number of critical biological pathways and functions: Here, we have mapped out all the pathways associated where GSK3β is the direct component. Among these pathways, a number of them influence human diseases directly, namely malignancies and infectious diseases, as well as disorders connected with the immune system. Wnt signaling, cell cycle chemokine signaling, BCR/TCR signaling, PI3K-AKT signaling, and insulin signaling believed to be shared by the illnesses type 2 diabetes and melanoma. The GSK3 interactors, type-2 diabetes, and cancer all have genes and pathways in common, whereas T2D and melanoma did not. In the case of melanoma, CDKN2A and TP53 might be the main medicines addressed by Diosmin and Fisetin, respectively.

Author Contributions: Conceptualization, D.A., and N.H.; methodology, D.A., and N.H.; software, D.A., and N.H.; validation, D.A., and N.H.; formal analysis, D.A., and N.H.; investigation, D.A., and N.H.; resources, D.A., and N.H.; data curation, D.A., and N.H.; writing—original draft preparation, D.A., and N.H.; writing—review and editing, D.A., and N.H.; visualization D.A., and N.H.; supervision, D.A., and N.H.; project administration, N.H.; funding acquisition, D.A., and N.H. All authors have read and agreed to the published version of the manuscript.

Funding: This research received no external funding.

Acknowledgments: We are thankful to DSR, KAU, Saudi Arabia and for providing us the resources and the facility to carry out the work.

Availability of data and materials: Not Applicable.

Dec 2024

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: All the related data are supplied in this work or have been referenced properly.

References

- 1. Beurel, E., S.F. Grieco, and R.S. Jope, *Glycogen synthase kinase-3 (GSK3): regulation, actions, and diseases.* Pharmacol Ther, 2015. **148**: p. 114-31.
- 2. Biondi, R.M. and A.R. Nebreda, Signalling specificity of Ser/Thr protein kinases through docking-site-mediated interactions. Biochemical Journal, 2003. **372**(1): p. 1-13.
- 3. Jope, R.S., C.J. Yuskaitis, and E. Beurel, *Glycogen synthase kinase-3 (GSK3): inflammation, diseases, and therapeutics.* Neurochem Res, 2007. **32**(4-5): p. 577-95.
- 4. Mobashir, M., et al., An Approach for Systems-Level Understanding of Prostate Cancer from High-Throughput Data Integration to Pathway Modeling and Simulation. Cells, 2022. **11**(24): p. 4121.
- 5. Maqbool, M., M. Mobashir, and N. Hoda, *Pivotal role of glycogen synthase kinase-3: A therapeutic target for Alzheimer's disease*. European Journal of Medicinal Chemistry, 2016. **107**: p. 63-81.
- 6. Schlicker, A., et al., Subtypes of primary colorectal tumors correlate with response to targeted treatment in colorectal cell lines. BMC Medical Genomics, 2012. **5**(1): p. 66.
- 7. Sun, Y., et al., MST2 methylation by PRMT5 inhibits Hippo signaling and promotes pancreatic cancer progression. EMBO J, 2023: p. e114558.
- 8. Manning, B.D. and L.C. Cantley, AKT/PKB Signaling: Navigating Downstream. Cell, 2007. 129(7): p. 1261-1274.
- 9. Suzuki, T., et al., Inhibition of AMPK Catabolic Action by GSK3. Molecular Cell, 2013. 50(3): p. 407-419.
- 10. Hornberg, J.J., et al., Cancer: A Systems Biology disease. Biosystems, 2006. 83(2-3): p. 81-90.
- 11. Mukherjee, N., et al., A Systems Biology Approach to Investigate Kinase Signal Transduction Networks That Are Involved in Triple Negative Breast Cancer Resistance to Cisplatin. J Pers Med, 2022. **12**(8).
- 12. Werner, H.M.J., G.B. Mills, and P.T. Ram, *Cancer Systems Biology: a peek into the future of patient care?* Nature Reviews Clinical Oncology, 2014. **11**(3): p. 167-176.
- 13. Elkins, J.M., et al., Comprehensive characterization of the Published Kinase Inhibitor Set. Nature Biotechnology, 2016. 34(1): p. 95-103.
- 14. Huber, K.V.M., et al., Stereospecific targeting of MTH1 by (S)-crizotinib as an anticancer strategy. Nature, 2014. 508(7495): p. 222-227.
- 15. Baba, M.R. and S.A. Buch, *Revisiting Cancer Cachexia: Pathogenesis, Diagnosis, and Current Treatment Approaches.* Asia-Pacific Journal of Oncology Nursing, 2021. 8(5): p. 508-518.
- 16. Muellner, M.K., et al., *A chemical-genetic screen reveals a mechanism of resistance to PI3K inhibitors in cancer.* Nature Chemical Biology, 2011. **7**(11): p. 787-793.
- 17. Chitforoushzadeh, Z., et al., *TNF-insulin crosstalk at the transcription factor GATA6 is revealed by a model that links signaling and transcriptomic data tensors.* Science Signaling, 2016. **9**(431): p. ra59.
- 18. Hilioti, Z., et al., GSK-3 kinases enhance calcineurin signaling by phosphorylation of RCNs. Genes & Development, 2004. 18(1): p. 35-47.
- 19. Klingmuller, U., et al., *Primary mouse hepatocytes for systems biology approaches: a standardized in vitro system for modelling of signal transduction pathways.* Syst Biol (Stevenage), 2006. **153**(6): p. 433-47.
- 20. Mobashir, M., et al., An Approach for Systems-Level Understanding of Prostate Cancer from High-Throughput Data Integration to Pathway Modeling and Simulation. Cells, 2022. **11**(24).
- 21. Varjosalo, M., et al., Application of Active and Kinase-Deficient Kinome Collection for Identification of Kinases Regulating Hedgehog Signaling. Cell, 2008. **133**(3): p. 537-548.
- 22. Zhou, A., et al., *Nuclear GSK3β promotes tumorigenesis by phosphorylating KDM1A and inducing its deubiquitylation by USP22.* Nature Cell Biology, 2016. **18**(9): p. 954-966.
- 23. Ahmed, S., et al., A Network-Guided Approach to Discover Phytochemical-Based Anticancer Therapy: Targeting MARK4 for Hepatocellular Carcinoma. Front Oncol, 2022. **12**: p. 914032.
- 24. Almowallad, S., L.S. Alqahtani, and M. Mobashir, *NF-kB in Signaling Patterns and Its Temporal Dynamics Encode/Decode Human Diseases.* Life (Basel), 2022. **12**(12).
- Almowallad, S., R. Jeet, and M. Mobashir, Systems-level understanding of toxicology and cardiovascular system. Jour. Bas. Sci., 2024.
 5(1): p. 1-16.
- 26. Anwer, S.T., et al., Synthesis of Silver Nano Particles Using Myricetin and the In-Vitro Assessment of Anti-Colorectal Cancer Activity: In-Silico Integration. Int J Mol Sci, 2022. 23(19).

- 27. Bajrai, L.H., et al., *Gene Expression Profiling of Early Acute Febrile Stage of Dengue Infection and Its Comparative Analysis With Streptococcus pneumoniae Infection.* Front Cell Infect Microbiol, 2021. **11**: p. 707905.
- 28. Bajrai, L.H., et al., Understanding the role of potential pathways and its components including hypoxia and immune system in case of oral cancer. Sci Rep, 2021. **11**(1): p. 19576.
- 29. Choudhry, H., et al., Study of APOBEC3B focused breast cancer pathways and the clinical relevance. Jour. Bas. Sci., 2024. 2(1): p. 1-12.
- 30. Eldakhakhny, B.M., et al., *In-Silico Study of Immune System Associated Genes in Case of Type-2 Diabetes With Insulin Action and Resistance, and/or Obesity.* Front Endocrinol (Lausanne), 2021. **12**: p. 641888.
- Huang, L., L. Liao, and C.H. Wu, Protein-protein interaction prediction based on multiple kernels and partial network with linear programming. BMC Systems Biology, 2016. 10(Suppl 2): p. 45.
- 32. Bigler, J., et al., Cross-study homogeneity of psoriasis gene expression in skin across a large expression range. PLoS One, 2013. 8(1): p. e52242.
- 33. Yang, X., et al., *Microarray profiling of skeletal muscle tissues from equally obese, non-diabetic insulin-sensitive and insulin-resistant Pima Indians*. Diabetologia, 2002. **45**(11): p. 1584-93.
- 34. Almowallad, S., R. Jeet, and M. Mobashir, *A systems pharmacology approach for targeted study of potential inflammatory pathways and their genes in atherosclerosis.* Jour. Bas. Sci., 2024. **6**(1): p. 1-12.
- 35. Jeet, R., *Phytomedicine as the alternative drug against the human disease*. Journal of Basic Science, 2024. 7(1).
- 36. Athar, M.T., Possible herbal medications and human cancer targets. Jour. Bas. Sci., 2025. 1(2): p. 1-12.
- 37. Huwait, E. and M. Mobashir, Potential and Therapeutic Roles of Diosmin in Human Diseases. Biomedicines, 2022. 10(5).
- 38. Brunk, E., et al., Systems biology of the structural proteome. BMC Systems Biology, 2016. 10(1): p. 26.
- 39. Helmi, N., D. Alammari, and M. Mobashir, *Role of Potential COVID-19 Immune System Associated Genes and the Potential Pathways Linkage with Type-2 Diabetes.* Comb Chem High Throughput Screen, 2022. **25**(14): p. 2452-2462.
- 40. Krishnamoorthy, P.K.P., et al., *In-silico study reveals immunological signaling pathways, their genes, and potential herbal drug targets in ovarian cancer.* Informatics in Medicine Unlocked, 2020. **20**: p. 100422.
- 41. El-Kafrawy, S.A., et al., Genomic profiling and network-level understanding uncover the potential genes and the pathways in hepatocellular carcinoma. Front Genet, 2022. **13**: p. 880440.
- 42. Kamal, M.A., et al., *Gene expression profiling and clinical relevance unravel the role hypoxia and immune signaling genes and pathways in breast cancer: Role of hypoxia and immune signaling genes in breast cancer.* Journal of Internal Medicine: Science & Art, 2020. **1**.
- 43. Khan, B., et al., Deciphering molecular landscape of breast cancer progression and insights from functional genomics and therapeutic explorations followed by in vitro validation. Scientific Reports, 2024. **14**(1).
- 44. Khouja, H.I., et al., *Multi-staged gene expression profiling reveals potential genes and the critical pathways in kidney cancer.* Sci Rep, 2022. **12**(1): p. 7240.
- 45. Warsi, M.K., et al., Comparative Study of Gene Expression Profiling Unravels Functions Associated with Pathogenesis of Dengue Infection. Curr Pharm Des, 2020. **26**(41): p. 5293-5299.
- 46. Pawson, T. and M. Kofler, *Kinome signaling through regulated protein–protein interactions in normal and cancer cells*. Current Opinion in Cell Biology, 2009. **21**(2): p. 147-153.
- 47. Song, S., et al., miR-3200 accelerates the growth of liver cancer cells by enhancing Rab7A. Noncoding RNA Res, 2023. 8(4): p. 675-685.
- 48. Biber, K., et al., *Central nervous system myeloid cells as drug targets: current status and translational challenges.* Nature Reviews Drug Discovery, 2016. **15**(2): p. 110-124.
- 49. Bringuier, C.M., et al., Up-Regulation of Astrocytic Fgfr4 Expression in Adult Mice after Spinal Cord Injury. Cells, 2023. 12(4).
- 50. Calderone, A., et al., *Comparing Alzheimer's and Parkinson's diseases networks using graph communities structure*. BMC Systems Biology, 2016. **10**(1): p. 25.
- 51. Coleman, M. and R. Ribchester, *Programmed Axon Death, Synaptic Dysfunction and the Ubiquitin Proteasome System.* Current Drug Target -CNS & Neurological Disorders, 2004. **3**(3): p. 227-238.
- 52. DiMauro, S. and E.A. Schon, Mitochondrial Disorders in the Nervous System. Annual Review of Neuroscience, 2008. 31(1): p. 91-123.
- 53. Rajagopal, V., et al., *An atlas of RNA-dependent proteins in cell division reveals the riboregulation of mitotic protein-protein interactions.* Nat Commun, 2025. **16**(1): p. 2325.
- 54. Szyf, M., Prospects for the development of epigenetic drugs for CNS conditions. Nature Reviews Drug Discovery, 2015. 14(7): p. 461-474.
- 55. Tan, L., J.-T. Yu, and L. Tan, *The kynurenine pathway in neurodegenerative diseases: Mechanistic and therapeutic considerations.* Journal of the Neurological Sciences, 2012. **323**(1-2): p. 1-8.

Disclaimer/Publisher's Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of Global Journal of Basic Science and/or the editor(s). Global Journal of Basic Science and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.

Copyright: © 2024 by the authors. Submitted for possible open access publication under the terms and conditions of the Creative Commons Attribution (CC BY) license (https://creativecommons.org/licenses/by/4.0/).